α-Select **Chemically Competent Cells**

Shipping: On Dry Ice

Catalog numbers

Batch No .: See vial BIO-85025 ≥10⁷ cfu/µg of pUC19 (Bronze Efficiency)

BIO-85026 ≥108 cfu/µg of pUC19 (Silver Efficiency)

BIO-85027 ≥109 cfu/µg of pUC19 (Gold Efficiency)



Store at -80°C

α-Select Chemically Competent Cells are shipped on Dry/Blue Ice and stored at -80°C.

When stored under the recommended conditions and handled correctly, full activity of the cells is retained until the expiry date on the outer box label.

Product Specifications:

Storage and stability

Control Vector Efficiency Pack Size Bronze 2ml (10 x 200µl) pUC19 (100pg/µl) Silver 2ml (10 x 200µl) pUC19 (10pg/µl)

1ml (20 x 50µl) pUC19 (10pg/µl) Genotype

 $F^{-} deo R end A1 \ rec A1 \ rel A1 \ gyr A96 \ hsd R17 (r_{k}^{-}, \ m_{k}^{+}) \ sup E44 \ thi-1 \ pho A \ \Delta (lac ZYA \ arg F) U169 \ \Phi 80 lac Z \Delta M15 A \ A10 \ A1$

Safety precautions

This product is for R&D use only, not for human use, or any other use. Please refer to the material safety data sheet for information regarding hazards and safe handling practice.

Notes

Research Use Only.

Features

- Chemically Competent or Electroporation Grade
- Variety of efficiencies: ≥10⁷, ≥10⁸, or ≥10⁹ cfu/µg of pUC19
- Accommodates larger plasmids

Applications

- Transformation of cloned DNA into bacterial cells
- Ideal for subcloning and generating cDNA libraries
- Blue/white color screening

Description

α-Select Competent Cells contain a lacZ marker that provides α -complementation of the β-galactosidase gene for blue/white color screening. The cells are ideal for generating cDNA libraries and subcloning.

α-Select Competent Cells also provide recA1 and endA1 markers to minimize recombination and enhance the quality of the plasmid DNA. pUC19 DNA is also provided as a positive control.

Suggested Transformation Procedure for Optimal Results:

- Remove cells from -80°C and let thaw on wet ice.
- Gently mix cells by lightly flicking tube. Aliquot ~50-100µl of cells into chilled, 17 x 100mm polypropylene tube(s), e.g., Falcon 2059. Unused cells may be refrozen, but a small drop in efficiency may result. For optimal recovery, refreeze cells in a dry ice/ ethanol bath prior to -80°C storage.
- Add DNA solution (≤5µl per 50µl cells) to cell suspension and gently swirl tube(s) for a few seconds to mix. If a control is desired, repeat this step with 2µl of the provided Control Vector (pUC19) in a separate
- 4. Incubate on ice for 30 minutes.
- Place tube(s) in 42°C water bath for ~30 to 45 seconds without shaking. For 50µl aliquots in Falcon 2059 tubes, 30 seconds is recommended for maximum efficiency.
- Replace tube(s) on ice for ~2 minutes.
- Dilute transformation reaction(s) to 1ml by addition of 900-950µl SOC. SOC Medium: 2% Tryptone, 0.5% Yeast Extract, 0.4% glucose, 10mM NaCl, 2.5mM KCl, 10mM MgCl₂ & 10mM MgSO₄.
- 8. Shake tube(s) ~200 rpm for 60 minutes at 37°C.
- Plate by spreading 5-200µl of cell transformation mixture on LB agar 9. plates containing appropriate antibiotic and incubate overnight at

When performing the pUC19 control transformation, plate 5µl of the transformation mixture on a LB agar plate containing 100µg/ml ampicillin. To facilitate cell spreading, place a pool of SOC (100µl) onto surface of plate prior to addition of transformation mixture.

Transformation Efficiency Calculation for Control Vector

Transformation Efficiency (cfu/μg pUC19 DNA)

colonies (colony forming units) pg pUC19

10⁶ pg μg

Final volume (μ I) of transformation mix

For example:

If 40 colonies were obtained after transforming 20pg of pUC19 and plating 5µl of the final 1ml transformation mixture, the calculated transformation efficiency would be:

20pg pUC19

10⁶ pg μg

5ul

4 x 108 cfu/μg pUC19

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Associated Products:

Product Name	Pack Size	Cat No
T4 DNA Ligase	500 Units	BIO-27026
Quick-Stick Ligase	50 Reactions	BIO-27027
IPTG	5g	BIO-37036
X-GAL	1g	BIO-37035

Product Citations:

- Zane, G. M., et al. Appl. Environ. Microbiol. 76(16), 5500-09
- Hornsey, M., et al. J. Antimicrob. Chemother. 65 (8), 1589-1593 2. (2010).
- Broeham, G., et al. Insect Biochem. Mol. Biol. 40(3), 274-283 (2010).
- Goldfinch, N., et al. Vet. Res. 41(5), 62 (2010).
- Thaler, A. D., et al. Conservation Gene. Res. DOI: 10.1007/s12686 -010-9174-9 (2010).
- Allerston, C.K., et al. Mol. Gene. Metab. 98(1-2), 198-202 (2009).

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